

# **PEPTONE WATER (7365)**

## Intended Use

**Peptone Water** is used for the cultivation of non-fastidious microorganisms, indole testing, and as a basal medium for carbohydrate fermentation studies.

#### **Product Summary and Explanation**

Peptone Water is a minimal growth medium. The formulation of Peptone Water permits cultivation of non-fastidious organisms.<sup>1</sup> This non-selective medium has been used as a basal medium for biochemical tests such as carbohydrate fermentation patterns and production of indole.<sup>1,2</sup> A 1% final carbohydrate concentration with phenol red (0.018 g/L) indicator and a Durham tube can be used for fermentation studies.

## Principles of the Procedure

Peptone Water contains Peptone as a source of carbon, nitrogen, vitamins, and minerals. Sodium Chloride maintains the osmotic balance.

## Formula / Liter

Peptone	10 a
Sodium Chloride	
Final pH: 7.2 ± 0.2 at 25°C	0
Formula may be adjusted and/or supplemented as required to meet r	oorformanco coocifi

Formula may be adjusted and/or supplemented as required to meet performance specifications.

## **Precautions**

- 1. For Laboratory Use.
- 2. IRRITANT. Irritating to eyes, respiratory system, and skin.

#### **Directions**

- 1. Dissolve 15 g of the medium in one liter of purified water.
- 2. Heat with frequent agitation to obtain solution.
- 3. Autoclave at 121°C for 15 minutes.

#### If Determining Carbohydrate Fermentation Patterns

- 1. Add 1.8 mL of a 1% phenol red solution to 1 liter rehydrated Peptone Water. Mix thoroughly.
- 2. Dispense into test tubes containing inverted Durham tubes.
- 3. Autoclave at 121°C for 15 minutes.
- 4. Aseptically add a sufficient sterile carbohydrate solution to yield a 1% final concentration. Rotate each tube thoroughly to distribute the carbohydrate.

#### **Quality Control Specifications**

**Dehydrated Appearance:** Powder is homogeneous, free flowing, and tan.

**Prepared Appearance:** Prepared medium is gold to amber, clear to trace hazy, and may have none to light ppt.

**Expected Cultural Response:** Cultural response in Peptone Water after 24 hours incubation at 35°C.

Microorganism	Growth Response w/o Carboydrates
Escherichia coli ATCC® 25922	growth
Salmonella typhimurium ATCC® 14028	growth
Staphylococcus aureus ATCC® 25923	growth



The organisms listed are the minimum that should be used for quality control testing.

## **Test Procedure**

## **Carbohydrate Fermentation**

Inoculate tubes with test organism. Incubate tubes at  $35 \pm 2^{\circ}$ C for 18 - 48 hours. Observe for color change.

#### Indole Test

Using aseptic technique, suspend the commercially available Indole Test Strip 100 mm above the surface of a 24 or 48 hour culture. Incubate at 37°C for 5 - 30 minutes.

## **Results**

#### Carbohydrate Fermentation Patterns

Acid is produced when carbohydrates are fermented. This is indicated by a yellow color in the medium. Gas production is indicated by the presence of gas bubbles in the Durham tube.

## **Indole Test**

Observe for the formation of a violet color on the strip indicating a positive test for indole production.

## **Storage**

Store sealed bottle containing the dehydrated medium at 2 - 30°C. Once opened and recapped, place container in a low humidity environment at the same storage temperature. Protect from moisture and light by keeping container tightly closed.

## Expiration

Refer to expiration date stamped on the container. The dehydrated medium should be discarded if not free flowing, or if appearance has changed from the original color. Expiry applies to medium in its intact container when stored as directed.

#### Limitation of the Procedure

Due to varying nutritional requirements, some strains may be encountered that grow poorly or fail to grow on this medium.

Packaging			
Peptone Water	Code No.	7365A	500 g
		7365B	2 kg
		7365C	10 kg

#### **References**

- 1. MacFaddin, J. F. 1985. Media for isolation-cultivation-identification-maintenance of medical bacteria, vol. 1, p. 610-612. Williams & Wilkins, Baltimore, MD.
- Balows, A., W. J. Hausler, K. L. Herrmann, H. D. Isenberg, and H. J. Shadomy (eds.). 1991. Manual of clinical microbiology, 5<sup>th</sup> ed. American Society for Microbiology, Washington, D.C.
- 3. Finegold, S. M., and W. Martin. 1982. Bailey and Scott's diagnostic microbiology, 6<sup>th</sup> ed. St. Louis.

#### Technical Information

Contact Acumedia Manufacturers, Inc. for Technical Service or questions involving dehydrated culture media preparation or performance at (517)372-9200 or fax us at (517)372-2006.

