XLD (Xylose Lysine Deoxycholate) Agar

Medium proposed by TAYLOR (1965), TAYLOR and HARRIS (1965, 1967) and TAYLOR and SCHELHART (1967) for the isolation and differentiation of pathogenic Enterobacteriaceae, especially of Shigella and Salmonella species.

This culture medium complies with the recommendations of the ISO 6579.

Mode of Action

Degradation of xylose, lactose and sucrose to acid causes phenol red to change its colour to yellow. Production of hydrogen sulfide is indicated by thiosulfate and iron(III) salt, which react to form a precipitate of black iron sulfide in the colonies. Bacteria which decarboxylate lysine to cadaverine can be recognized by the appearance of a purple colouration around the colonies due to an increase in pH.

These reactions can proceed simultaneously or successively, this may cause the pH indicator to exhibit various shades of colour or it may change its colour from yellow to red on prolonged incubation. The culture medium is weakly inhibitory.

Typical Composition (g/litre)

Yeast extract 3.0; sodium chloride 5.0; D(+)xylose 3.75; lactose 7.5; sucrose 7.5; L(+)lysine 5.0; sodium deoxycholate 1.0; sodium thiosulfate 6.8; ammonium iron(III) citrate 0.8; phenol red 0.08; agar-agar 14.5.

Preparation

- 1. Weigh out 55 g of XLD Agar.
- 2. Add 50 ml of demin. water to a flask
- 3. Transfer 55 g of XLD Agar gently to flask with swirling.
- Mix thoroughly, add remaining 950 ml demin. water, until completely suspended. Check for lumps. If present repeat mixing.
- 5. Heat to boiling to dissolve completely.
- 6. Immediately cool the medium to about 47-50 °C in a waterbath set at this temperature. Agitate flask to cool rapidly.
- 7. Pour plates.
- 8. Dry plates and check for sterility prior to use.

Note: preparation of large volumes, overheating and prolonged storage in water bath (47-50 °C) should be avoided.

Do not autoclave.

pH: 7.4 ± 0.2 at 25 °C.

The plates are clear and red.

Crystalline precipitate of salts may occur. To avoid this, the liquid medium needs to be filtered through a flute-formed filter.

Experimental Procedure and Evaluation

Inoculate by spreading the material thinly on the surface of the plates.

Incubation: up to 48 hours at 35 °C aerobically.

Further tests should be performed in order to identify the colonies.

Appearance of Colonies	Microorganisms		
Yellow, surrounded by yellow zones, opaque with precipitation zones	Escherichia coli, Enterobacter, Aeromonas		
Yellow, surrounded by yellow zones, opaque, mucoid with pre- cipitation zones	Klebsiella		
Yellow, surrounded by yellow zones, opaque, sometimes with a black centre	Citrobacter (lactose-positive strains)		
Yellow, surrounded by yellow zones, opaque,	Serratia, Hafnia		
Yellow, surrounded by yellow zones, translucent, black centre	Proteus vulgaris, most Pro- teus mirabilis		
Colonies have the same colour as the culture medium, translucent, sometimes with a black centre	Salmonella		
Colonies have the same colour as the culture medium, translucent	Shigella, Providencia, Pseudomonas		
Orange, slightly opaque	Salmonella typhosa (xylose-positive strains)		

Literature

American Public Health Association. Compendium of Methods for the microbiological Examination of Foods. – 3rd ed. (1992).

BHAT, P., a. RAIAN, D.: Comparative evaluation of deoxycholate citrate medium and xylose lysine deoxycholate medium in the isolation of shigellae. – Am. J. Clin. Pathol., 64; 99-404 (1975).

DUNN, C., a. MARTIN, W.J.: Comparison of media for isolation of Salmonellae and Shigellae from fecal specimens. – Appl. Microbiol., 22; 17-22 (1971). European Pharmacopeia II, Chapter VIII, 10.

ROLLENDER, W., BECKFORD, O., BELSKY, R.D., a. KOSTROFF, B.: Comparison of xylose lysine deoxycholate agar and MacCONKEY Agar for the isolation of Salmonella and Shigella from clininal specimens. – Am. J. Clin. Pathol., 51/2; 284-386 (1969).

TAYLOR, W.J.: Isolation of Shigellae. I. Xylose lysine agars: new media for isolation of enteric pathogens. – Am. J. Clin. Path., 44; 471-475 (1965).

TAYLOR, W.J., a. HARRIS, B.: Isolation of Shigellae. II. Comparison of plating media and enrichment broths. – Am. J. Clin. Path., 44X 476-479 (1965). TAYLOR, W.J., a. HARRIS, B.: Isolation of Shigellae. III. Comparison of new and traditional media with stool specimens. – Amer. J. Clin. Pathol., 48; 350-355 (1967).

TAYLOR, W.J., a. SCHELHART, D.: Isolation of Shigellae. IV. Comparison of plating media with stools. – Amer. J. Clin. Pathol., 48; 356-362 (1967). TAYLOR, W.J., a. SCHELHART, D.: Isolation of Shigellae. V. Comparison of enrichment broth with stools. – Appl. Microbiol., 16; 1383-1386 (1968). United States Pharmacopeia XXVI, Chapter "Microbioal Limit Tests", 2003

XLD (Xylose Lysine Deoxycholate) Agar

Ordering Information

Product	Merck Cat. No.	Pack size
XLD (Xylose Lysine Deoxycholate) Agar	1.05287.0500	500 g

Quality control (spiral plating method)

Test strains	lnoculum (cfu/ml)	Recovery rate %	Colony colour	Black centre	Colour change of medium
Escherichia coli ATCC 25922	> 10 ⁵	none to poor	yellow	-	yellow + precipitate
Enterobacter cloacae ATCC 13047	10 ³ -10 ⁵	≥ 30	yellow	-	yellow + precipitate
Klebsiella pneumoniae ATCC 13883	10 ³ -10 ⁵	≥ 30	yellow	-	yellow + precipitate
Shigella flexneri ATCC 12022	10 ³ -10 ⁵	≥ 10	colourless	-	
Shigella sonnei ATCC 11060	10 ³ -10 ⁵	≥ 10	colourless	-	
Salmonella typhimurium ATCC 14028	10 ³ -10 ⁵	≥ 30	colourless	+	-
Salmonella enteritidis NCTC 5188	10 ³ -10 ⁵	≥ 30	colourless	+	-
Proteus mirabilis ATCC 14273	10 ³ -10 ⁵	≥ 30	yellow	+	yellow / orange
Enterococcus faecalis ATCC 11700	> 10 ⁵	none		-	



Klebsiella pneumoniae ATCC 13883



Salmonella enteritidis NCTC 5188